

Original Research Article

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Varying Drug Resistant Patterns of MRSA Isolates with PVL Gene

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A B S T R A C T

Nosocomial infections due to Methicillin Resistant Staphylococcus aureus (MRSA) has been an element of concern to the medical personnel for the past four decades. Community associated MRSA (CA-MRSA) which was initially considered as more sensitive than hospital acquired MRSA (HA-MRSA) is now presenting with increasing levels of drug resistance. Along with the *mecA* gene, *pvl* gene is characteristically present in most isolates of CA-MRSA. To study the varying drug resistant patterns of MRSA isolates with *pvl* gene. A total of 150 clinical isolates of MRSA analysed in the study were subjected to susceptibility testing to cefoxitin (30 µg) and growth on oxacillin screen agar containing 6 µg/mL of oxacillin for the detection of methicillin resistance. All the isolates which were included in the study, were checked by PCR for the presence of *mecA* gene, which codes for methicillin resistance and for *pvl* gene. Amplification of 540bp and 625bp gene fragments in the PCR reaction indicates the presence of *mecA* and *pvl* genes respectively. *mecA* gene was present in all the 150 isolates of MRSA and *pvl* gene was present only in 26 isolates. Of these 26 isolates that had *pvl* gene, 14 were in MRSA isolated from outpatient samples and were sensitive to most of the non- beta lactam antibiotics. Among the other 12 inpatient MRSA isolates which had the *pvl* gene, nine were sensitive to most of the non- beta lactam antibiotics, whereas remaining three were resistant to most of the antibiotics except vancomycin and linezolid. The presence of *pvl* gene can no longer be used to discriminate between CA-MRSA and HA-MRSA. Indiscriminate empirical treatment of MRSA infections with high end antibiotics like glycopeptides needs to avoided and therapy with non beta lactam antibiotics like lincosamides which have better soft tissue penetration should be used as very few new antimicrobial agents are in the pipeline.

Keywords

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Introduction

Traditionally MRSA has been considered as a major nosocomial pathogen in health care facilities, but in the last decade it has also been observed to be an emerging pathogen causing increasing number of community acquired infections. The emergence of this pathogen depends on its ability to survive in

different environments and to interact successfully with the host. It was initially presumed that community acquired MRSA infection is attributed to the presence of Panton Valantine Leucocidin (PVL) toxin encoded by *pvl* gene. However, the presence of *pvl* gene alone cannot be considered as a

marker of CA-MRSA (Vandenesch *et al.*, 2003; Shenoy *et al.*, 2010). The community acquired strains tend to be more susceptible to non- beta lactam agents as compared to hospital acquired MRSA isolates and appear to carry a unique Staphylococcal chromosome (SCC mec type IV) in relation to resistant genes.

MRSA are highly virulent strains capable of clonal dissemination and have the ability to cause epidemics of furunculosis and other skin and soft tissue infections, irrespective of characteristics of populations or the health care setting (Rachel, 2008; Harbarth *et al.*, 2005). The MRSA isolates carrying the *PVL* toxin possesses a serious threat and is a major public health concern (Vandenesch *et al.*, 2003; Shenoy *et al.*, 2010). The aim of the present study was to describe the varying drug resistant patterns of MRSA isolates with *pvl* gene.

Methods

This prospective study was carried out in the Department of Clinical Microbiology, Pondicherry Institute of Medical Sciences (PIMS). All the clinically significant 150 MRSA isolates were included in the study.

The clinical isolates were screened for methicillin resistance by susceptibility to cefoxitin (30 µg) and growth on oxacillin screen agar containing 6 µg /mL of oxacillin. Antibiotic susceptibility testing was done on Muller Hinton agar by Kirby Bauer disc diffusion method for the following antibiotics; Trimethoprin–Sulphamethoxazole (1.25/23.75 µg), Tetracycline (30µg), Linezolid (30 µg), Erythromycin (15µg), Clindamycin (2 µg), Teicoplanin (30 µg), Chloramphenicol (30 µg), Ciprofloxacin (5 µg), Penicillin (10 units), Amikacin (30 µg) and Gentamicin (10 µg) (Hi-media) were recorded as per CLSI guidelines. The MIC for vancomycin against the 150 MRSA Isolates was determined by agar dilution method.

Molecular Methods for MRSA detection

mecA gene, which codes for Methicillin resistance and *pvl* gene were detected using PCR for all the 150 MRSA isolates included in the study. The following primers procured from Hi-media (India) were used in the study.

mecA (forward primer 1 µL; 5' GTA GAA ATG ACT GAA CGT CCG ATAA 3'), (Reverse primer 1 µL; 5' CCA ATT CCA CAT TGT TTC GGT CTAA 3').

*Pvl*gene (forward primer 1µL; 5'ATCATTAGGTAAAATGTCTGGACATG ATCC A-3'), (Reverse primer 1 µL; 5'GCATC AST GTA TTG GAT AGC AAA AGC – 3').

The amplified products were analysed by agarose gel electrophoresis. Amplification of 540bp and 625bp gene fragments in the PCR reaction indicates the presence of *mecA* and *pvl* genes respectively. The positive control strain used in PCR for *mecA* was ATCC 43300 and the negative control was ATCC 25973.

Clinical details of the patient and the demographic information, history of antibiotic usage, previous hospitalization, co-morbidities like Diabetes, post surgical status were recorded in a designed proforma. The patients from whom MRSA was isolated were followed up and the responses to treatment were noted.

Results and Discussion

A total of 354 *Staphylococcus aureus* were isolated out of which 150 were found to be MRSA. Rate of methicillin resistance among the *Staphylococcus aureus* isolates was 42.37%. Among the 150 patients from whom MRSA was isolated, 68 patients (45.67%) had a prolonged hospital stay of greater than 7

days duration, 62 patients (41.25%) had surgical interventions, 59 patients (39.6%) were referred from other hospitals and 39 patients (26.1%) were admitted in ICU for more than 2 days. Isolates from exudates which included pus, wound swab, and sterile body fluids (apart from blood) constituted 90.67 % (n=136) of isolates. Clinically significant respiratory isolates were from tracheal aspirate and bronchoalveolar lavage which was found to be 7.3% (n = 11). Two isolates were from blood and only one isolate was from urine (Figure1).

mecA gene was present in all the 150 isolates of MRSA and *pvl* gene was present only in 26 isolates. All the 26 MRSA isolates with *pvl* gene were from patients with skin and soft tissue infection. The MRSA isolates from blood and other sterile sites did not carry the *pvl* gene. Of the 26 isolates which had *pvl* gene, 14 were from MRSA isolated from outpatient samples and were sensitive to most of the non- beta lactam antibiotics and 12 were from MRSA isolated from inpatient samples. Among these 12 inpatient MRSA isolates which had the *pvl* gene, nine were sensitive to most of the non- beta lactam antibiotics, whereas remaining three isolates were resistant to the non beta lactam antibiotics like quinolones, macrolides, lincosamide and tetracycline except vancomycin and linezolid. Multidrug resistant MRSA strains were detected in the study. Only three isolates showed multidrug resistance, two of them exhibited inducible clindamycin resistance and one of them exhibited constitutive resistance to clindamycin.

Among these three isolates, one of them was from the wound swab of a patient with diabetic foot and the other two were from gluteal abscess pus and post auricular abscess. Following the culture report they were started on oral linezolid to which they responded well.

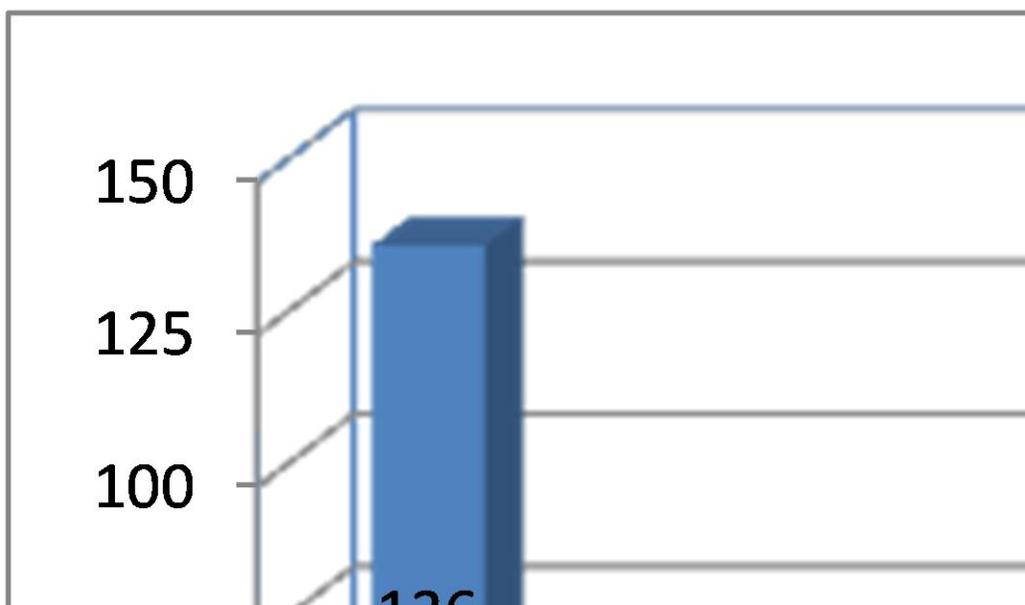
Historically, infections caused by methicillin-resistant *Staphylococcus aureus* (MRSA) were predominantly associated with patients in hospitals and skilled nursing facilities. In recent years, reports of community-associated MRSA infections (CA-MRSA) have been increasing (Vandenesch *et al.*, 2003; Shenoy *et al.*, 2010). Several studies conducted in India and elsewhere have shown that the prevalence of CA-MRSA to be varied ranging from 15 to 32% (Vandenesch *et al.*, 2003; Shenoy *et al.*, 2010). A significant difference has been demonstrated in the susceptibility of CA-MRSA and HA-MRSA by various investigators (Hacek *et al.*, 2009; Diep *et al.*, 2006; Ruhe *et al.*, 2007). Hospital acquired MRSA (HA-MRSA) have been found to exhibit resistance to frequently used non beta-lactam antibiotics whereas Community acquired MRSA (CA-MRSA) are known to cause skin and soft tissue infections and are usually susceptible to lincosamides like clindamycin which have better tissue penetration and absorption.

Diverse spectrum Studies by Ruhe and Menon have suggested that non beta-lactam antibiotics like tetracycline, which can be administered orally as a feasible option for treatment of CA-MRSA infection (Ruhe *et al.*, 2007). However, few strains of CA-MRSA have shown to exhibit multi drug resistance to most of the commonly used non beta lactam antibiotics. The incidence of CA-MRSA was 17.33% (n=26/150) in the present study which is in concurrence with the findings of various other studies. However, these investigators have studied the incidence of CA-MRSA in the community. The incidence of CA-MRSA in a tertiary care centre, as studied by Kanerva *et al.*, (2009) is found to be 21%. Most of the CA-MRSA isolates (88%) were primarily found to be susceptible to antibiotics like clindamycin, amikacin, erythromycin, cotrimoxazole, tetracycline and chloramphenicol, unlike HA-MRSA.

A study done by Nandita *et al.*, 2016 have shown an increased rate of susceptibility to clindamycin (84%) and have recommended the presence of *pvl* gene along with clindamycin susceptibility to be a predictor to detect community origin of MRSA (Nandita *et al.*, 2016). In our study also, it was found

that the 88.5% of isolates carrying the *pvl* gene and susceptible to clindamycin was found to be statistically significant ($p < 0.001$). Clindamycin has an excellent tissue penetration and is effective for the treatment of skin and soft tissue infections (Mandelia *et al.*, 2010).

Fig.1 Distribution of the isolates of MRSA



It has been reported that the action of *pvl* toxin in the tissue is decreased even by sub inhibitory concentration of clindamycin (Dumitrescu *et al.*, 2007). However, three among the 26 CA-MRSA isolates showed multi drug resistance and were sensitive only to vancomycin and linezolid. Among the MRSA isolates carrying the *pvl* gene, two of them exhibited inducible clindamycin resistance which is in contrast to Vysakh *et al.*, wherein they reported that 44.4% of their MRSA isolates carrying the *pvl* gene showed inducible clindamycin resistance (Vysakh *et al.*, 2013).

Emergence of multi drug resistance among CA-MRSA isolates is of major concern to the society. This is attributed to the indiscriminate empirical use of high end antibiotics like vancomycin and linezolid by many clinicians

without using other non beta lactum antibiotics as the first line of management. In our study, the prevalence of CA-MRSA with *pvl* gene positive isolates was 17.33 %. Studies on prevalence of *pvl* gene in various geographical settings have shown varied results, where the prevalence varies from 13.6 to 21.7 %. The presence of *pvl* gene can no longer be used to discriminate between CA-MRSA and HA-MRSA. However, MRSA isolates carrying the *pvl* gene are predominantly associated with skin and soft tissue infections. Clindamycin, which is a lincosamide has an excellent skin and soft tissue penetration and can be a potent choice to treat infections caused by strains of MRSA carrying the *pvl* gene.

Prevention of CA-MRSA is of utmost importance in health care setting ⁽¹⁴⁾. Simple

techniques like hand washing, barrier nursing and screening of patients for MRSA referred from other healthcare centres prior to admission for MRSA provide a long way for controlling the transmission of CA-MRSA infection.

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